LESSON PLAN

Name of the Faculty : Mr.Amit Kumar

Discipline : DMLT

Semester : 3rd(2nd Year)

Subject : HISTOPATHOLOGY

Lesson Plan Duration : 16 weeks(from 20 August 2024 to 29 November 2024)(According to Syllabus Scheme)

Work load(Lecture/Practical)per week(in hours): Lectures-03(hr),Practicals-04(hr)

Week	Theory		Practical		
	Lecture Day	Topic(including assigment/test)	Practical Day	Торіс	
	1st	Introduction and definition (1st Unit)			
1st	2nd	Reception of Specimen	1st	Reception of specimen, labeling and preserving the specimen	
	3rd	Various Terms associated with staining			
	1st	Metachromasia, Accelerators			
2nd	2nd	Progressive and regressive staining	2nd	Preparation of various smears by unfixed methods	
	3rd	Use of controls in staining and their significance			
	1st	Haematoxylin and Eosin staining			
3rd	2nd	Haematoxylin and Eosin staining	3rd	Preparation of different fixatives with special emphasis	
	3rd	Deparaffinization, Hydration, Nuclear Staining			
	1st	Differentiation, Blueing, Counterstaining,			
4th	2nd	Dehydration	4th	Preparation of paraffin blocks from various tissue pieces and labeling	
	3rd	Clearing/Dealcoholization			
	1st	Automation: Use of automatic stainer			
5th	2nd	automatic coverslipper	5th	Handling of microtome	
	3rd	Preparation of Tissue (Different Methods) (2nd Unit)			
	1st	Unfixed Tissue preparations			
6th	2nd	Fixed Tissue preparations	6th	Sharpening of microtome knives	
	3rd	Classification of fixatives (3rd Unit)			
	1st	Classification of fixatives			
7th	2nd	Composition of various fixatives	7th	Preparation of blocks for fine cutting	
	3rd	Processing of Tissue (by Paraffin Technique)			
	1st	Dehydration,Clearing/Dealcoholization			
8th	2nd	Infilteration and impregnation	8th	Practice of fine section cutting	
	3rd	Paraffin embedding			

	1st	Various types of mounting media		
9th	2nd	Advantages and Disadvantages	9th	Performing H&E staining on sections and mounting of tissue sections
	3rd Microtome, Types, Advantages and disadvantages	Microtome, Types, Advantages and disadvantages (4th Unit)		
	1st	Automation: Automatic knife sharpener		
10th	2nd	Various types of knives	10th	Demonstration of cell using buccal smear/urine sample
	3rd	Sharpening of knives		
	1st	Automation: Automatic knife sharpener		
11th	2nd	Uses of abrasives and lubricants	11th	Processing of urine samples for malignant cells
	3rd	Section Cutting		
	1st	Use of tissue floatation bath		
12th	2nd	Use of various adhesive media and lifting of sections to the slide	12th	Processing of sputum sample for malignant cytology
	3rd	Errors /cutting faults in sections and their remedies		
	1st	Exfoliative Cytology (5th Unit)		
13th	2nd	Preparation of vaginal & cervical smears	13th	To perform PAP stain on given smear
	3rd	Collection and Processing of specimen for cytology		
	1st	Collection and Processing of specimen for cytology		
14th	2nd	Role of cytotechnician in cytology	14th	To perform MGG stain on given smear
	3rd	Role of cytotechnician in cytology		
	1st	Papanicalaou staining		
15th	2nd	Papanicalaou staining	15th	To perform H&E on given smear
	3rd	May Grunwald & Giemsa staining		
	1st	May Grunwald & Giemsa staining		
16th	2nd	Sessional,Test	16th	Practical assignment
	3rd	Sessional,Test		

LESSON PLAN

Name of the Faculty Mrs. PARMILA Devi

Discipline DMLT

Semester 3rd(2nd year)

Subject PARASITOLOGY AND VIROLOGY

Lesson Plan Duration : 16 weeks(from 20 August 2024 to 29 November 2024)(According to Syllabus Scheme)

Work load(Lecture/Practical)per week(in hours): Lectures-03(hr) Practicals-04(hr)

Week		Theory		Practical
	Lecture Day	Topic(including assigment/test)	Practical Day	Topic
	1st	Introduction to medical parasitology (1st unit)		Collection and routing steel examination for detection of intestinal paracites using
1st	2nd	Introduction to medical parasitology	1st	Collection and routine stool examination for detection of intestinal parasites using
	3rd	General characteristics, morphology, classification		Saline preparation.
	1st	General characteristics, morphology, classification		Collection and routine stool examination for detection of intestinal parasites using
2nd	2nd	Laboratory Samples for detection of parasites (2nd unit)	2nd	Lugol's lodine preparation.
	3rd	Laboratory Samples for detection of parasites		Lugor 3 lourne preparation.
	1st	Concentration techniques		
3rd	2nd	Concentration techniques	3rd	Concentration methods of stool examination by Floatation method
	3rd	Concentration techniques		
	1st	Concentration techniques		Concentration methods of stool examination by Sedimentation methods
4th	2nd	Concentration techniques	4th	
	3rd	Concentration techniques		
	1st	Giardia and Entamoeba histolytica (3rd unit)		Identification of following adult worms/cyst from preserved specimen/slides
5th	2nd	Giardia and Entamoeba histolytica	5th	
	3rd	Giardia and Entamoeba histolytica		
	1st	Giardia and Entamoeba histolytica		
6th	2nd	Giardia and Entamoeba histolytica	6th	Identification of following adult worms/cyst from preserved specimen/slides
	3rd	Ancylostoma and Ascaris lumbricoides		
	1st	Ancylostoma and Ascaris lumbricoides		
7th	2nd	Ancylostoma and Ascaris lumbricoides	7th	Identification of following adult worms/cyst from preserved specimen/slides
	3rd	Ancylostoma and Ascaris lumbricoides		
	1st	Ancylostoma and Ascaris lumbricoides		
8th	2nd	Ancylostoma and Ascaris lumbricoides	8th	Identification of following adult worms/cyst from preserved specimen/slides
	3rd	T solium, T saginata		

	1st	T solium, T saginata		
9th	2nd	T solium, T saginata	9th	Identification of following adult worms/cyst from preserved specimen/slides
3(1)			- 501	identification of following addit worms/cyst from preserved specifien/slides
	3rd	T solium, T saginata		
	3rd	T solium, T saginata		
10th	1st	Malarial Parasite (P. Vivax and P. Falciparum) (4th unit)	10th	Preparation of smear and identification of blood parasites
	2nd	Malarial Parasite (P. Vivax and P. Falciparum)		
	3rd	Malarial Parasite (P. Vivax and P. Falciparum)		
11th	1st	Malarial Parasite (P. Vivax and P. Falciparum)	11th	Preparation of Leishman stain
	2nd	Malarial Parasite (P. Vivax and P. Falciparum)		
	3rd	Malarial Parasite (P. Vivax and P. Falciparum)		
12th	1st	Virology (5th unit)		Preparation of Giemsa stain
	2nd	Virology		
	1st	Virology		
13th	2nd	Virology	13th	Preparation of Field stain
	3rd	Medically important viruses		
	3rd	Medically important viruses	14th	
14th	1st	Medically important viruses		Preparation of thin and thick smears
	2nd	Medically important viruses		
	3rd	Virological Samples		
15th	1st	Virological Samples	15th	Staining of smears by Leishman, Giemsa, and Field stain.
	2nd	Sessional,Test		
	3rd			
16th	1st		16th	Examination of smears for malarial parasite (P. vivax and P. falciparum)
	2nd			

LESSON PLAN

Name of the Faculty : Mrs. PARMILA Devi

Discipline : DMLT

Semester : 3rd(2nd year)

Subject : TRANSFUSION MEDICINE(Blood Bank)

**Lesson Plan Duration**: 16 weeks(from 20 August 2024 to 29 November 2024)(According to Syllabus Scheme)

Work load(Lecture/Practical)per week(in hours): Lectures-03(hr), Practicals-02(hr)

Week		Theory			Practical
	Lecture Day	Topic(including assigment/test)	Practi	tical Day	Торіс
	1st	Historical introduction to Transfusion medicine (blood banking) (1st unit	t)		
1st	2nd	Historical introduction to Transfusion medicine (blood banking )	1	1st	Performing ABO blood grouping by following method
	3rd	Antigen and Antibody			
	1st	Antigen and Antibody			
2nd	2nd	Antigen and Antibody	2	2nd	Performing ABO blood grouping by following method
	3rd	ABO Blood Group System (2nd	l unit)		
	1st	ABO Blood Group System			
3rd	2nd	ABO Blood Group System	3	3rd	Performing ABO blood grouping by following method
	3rd	ABO Blood Group System			
	1st	The Rh Blood Group System			
4th	2nd	The Rh Blood Group System	4	4th	Performing-Rh grouping by following techniques
	3rd	The Rh Blood Group System			
	1st	The Rh Blood Group System			
5th	2nd	Anticoagulants used in blood bank (3rd u	<b>unit)</b> 5	5th	Performing-Rh grouping by following techniques
	3rd	Anticoagulants used in blood bank			
	1st	Anticoagulants used in blood bank			
6th	2nd	Anticoagulants used in blood bank	6	6th	Performance of Coombs Test
	3rd	·	unit)		
	1st	Criteria for selection of Donor			
7th		Blood Collection and storage	7	7th	Performance of Coombs Test
	3rd	Blood Collection and storage			
	1st	Blood Collection and storage			
8th	2nd	Blood Collection and storage	8	8th	Cross Matching (compatibility testing)
	3rd	Various blood components (Packed cells, Fresh frozen plasma			

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	1st	Various blood components (Packed cells, Fresh frozen plasma		
9th	2nd	Various blood components (Packed cells, Fresh frozen plasma	9th	Cross Matching (compatibility testing)
	3rd	Various blood components (Packed cells, Fresh frozen plasma		
	1st	Test for blood transfusion (5th unit)		
10th	2nd	Cross Matching	10th	Preparation of anticoagulants
	3rd	Cross Matching		
	1st	Cross Matching		
11th	2nd	Coombs Test	11th	Malarial Parasite test by Thick and Thin smear preparation
	3rd	Coombs Test		
	1st	Coombs Test		
12th	2nd	Coombs Test	12th	VDRL Test
	3rd	Blood Transfusion reactions		
	1st	Blood Transfusion reactions		
13th	2nd	Blood Transfusion reactions	13th	HIV Test
	3rd	Blood Transfusion reactions		
	1st	Blood Transfusion reactions		
14th	2nd	Blood Transfusion reactions	14th	HbsAg Test
	3rd	Sessional,Test		
	1st	Sessional,Test		
15th	2nd	Sessional,Test	15th	HCV Test
	3rd	Sessional,Test		
	1st			
16th	2nd		16th	Preparation of platelet rich plasma and platelet poor plasma
	3rd			

LESSON PLAN

Name of the Faculty : Mrs. Poonam

Discipline : DMLT

Semester : 3rd(2nd year)

**Subject** : Clinical Haematology-I

Lesson Plan Duration : 16 weeks(from 20 August, 2024 to 29 November, 2024)(According to Syllabus Scheme)

Work load(Lecture/Practical)per week(in hours): Lectures-03(hr),Practicals-04(hr)

Week	/eek Theory			Practical
	Lecture Day	Topic(including assigment/test)	Practical Day	Торіс
		ESR and PCV		
	1st	Introduction		
1st	2nd	Various methods of estimation of ESR and their merits and demerits	1st	Estimation of ESR by wintrobe method
	3rd	Various methods of estimation of ESR and their merits and demerits		
	1st	Various methods of estimation of ESR and their merits and demerits		
2nd	2nd	Various methods of estimation of PCV and their merits and demerits	2nd	Estimation of ESR by wintrobe method
	3rd	Various methods of estimation of PCV and their merits and demerits		
	1st	Various methods of estimation of PCV and their merits and demerits		Estimation of ESR by westergren method
3rd	2nd	Factors involved in ESR	3rd	
	3rd	Factors involved in ESR		
	1st	Interpretation of results		
4th	2nd	Interpretation of results	4th	Estimation of ESR by westergren method
	3rd	All Unit Revision		
		Red Cell Indicies- MCV, MCH, MCHC		
	1st	Definition, reference range, calculation and interpretation of MCV		
5th	2nd	Definition, reference range, calculation and interpretation of MCV	5th	Estimation of PCV by wintrobe method
	3rd	Definition, reference range, calculation and interpretation of MCH		
	1st	Definition, reference range, calculation and interpretation of MCH		
6th	2nd	Definition, reference range, calculation and interpretation of MCHC	6th	Estimation of PCV by Capillary method
<b>J</b>		Supravital stain and reticulocyte counting		Estimation of FeV by capitally method
	3rd	Introduction		

	1st	Principle and procedure of staining and calculation		
7th	2nd	Principle and procedure of staining and calculation	7th	Calculate Red Cell Indicies – MCV, MCH, MCHC
	3rd	Principle and procedure of staining and calculation		, ,
	1st	Reference values & interpretation		
8th	2nd	Variation in Physiological Values such as Hb, PCV	8th	Counting of Reticulocyte in blood
	3rd	Variation in Physiological Values such as T.L.C and Platelet count		
		Anemias		
	1st	Definition and classification		
9th	2nd	Definition and classification	9th	To perform red cell fragility test on blood
	3rd	Definition and classification		
	1st	Laboratory diagnosis of: Iron deficiency anaemia		
10th	2nd	Laboratory diagnosis of: Iron deficiency anaemia	10th	To perform Sickling test on blood
	3rd	Laboratory diagnosis of: Iron deficiency anaemia		
	1st	Laboratory diagnosis of: Megaloblastic anaemia		
11th	2nd	Laboratory diagnosis of: Megaloblastic anaemia	11th	Estimation of foetal haemoglobin by alkali denaturation test
	3rd	Laboratory diagnosis of: Haemolytic anaemia		
	1st	Laboratory diagnosis of: Haemolytic anaemia		Estimation of plasma haemoglobin
12th	2nd	Laboratory diagnosis of: Sickle cell anaemia	12th	
	3rd	Laboratory diagnosis of: Sickle cell anaemia		
	1st	Laboratory diagnosis of: Thallasseamia		Estimation of and G6PDby Methylene Blue Reduction Test
13th	2nd	Laboratory diagnosis of: Thallasseamia	13th	
	3rd	Aplastic anaemia		
	1st	Aplastic anaemia		
14th		Red Cell fragility test	14th	To perform red cell fragility test on blood.
	2nd	Principle and procedure		
	3rd	Principle and procedure		
4=.1	1st	Principle and procedure	45.1	
15th	2nd	Principle and procedure	15th	
	3rd	Principle and procedure		
4611	1st	Principle and procedure	461	
16th	2nd	Principle and procedure	16th	
	3rd	Clinical significance		

LESSON PLAN

Name of the Faculty : Mrs. Poonam Rani

Discipline : DMLT

Semester : 3rd(2nd year)

Subject : APPLIED CLINICAL BIOCHEMISTRY

**Lesson Plan Duration** : 16 weeks(from 20 August, 2024 to 29 November, 2024)(According to Syllabus Scheme)

Work load(Lecture/Practical)per week(in hours): Lectures-03(hr),Practicals-02(hr)

Week	Veek Theory			Practical
	Lecture Day	Topic(including assigment/test)	Practical Day	Topic
		Serum Bilirubin		
	1st	Formation of bile pigments		
1st	2nd	Formation and excretion of bilirubin	1st	Serum bilirubin estimation
	3rd	Formation and excretion of bilirubin		
	1st	Conjugated and unconjugated bilirubin		
2nd	2nd	Principle and procedures of serum bilirubin estimation (Direct & Indirect)	2nd	Serum bilirubin estimation
	3rd	Reference values & Clinical significance		
		SGOT and SGPT		
	1st	Principle and procedures of estimation		
3rd	2nd	Principle and procedures of estimation	3rd	SGOT estimation
	3rd	Principle and procedures of estimation		
	1st	Principle and procedures of estimation		
4th	2nd	Reference values & Clinical significance	4th	SGOT estimation
	3rd	Reference values & Clinical significance		
		ALP and ACP		
	1st	Principle and procedures of estimation		
5th	2nd	Principle and procedures of estimation	5th	SGPT estimation
	3rd	Principle and procedures of estimation		
	1st	Principle and procedures of estimation		
6th	2nd	Reference values & Clinical significance	6th	SGPT estimation
	3rd	Reference values & Clinical significance		
	1st	Principle and procedures of estimation Serum Amylase		
7th	2nd	Principle and procedures of estimation	7th	Serum amylase estimation
	3rd	Reference values & Clinical significance		

		Serum Calcium and Potassium		
	1st	Principle and procedures of estimation		
8th	2nd	Principle and procedures of estimation	8th	ALP estimation
	3rd	Principle and procedures of estimation		
	1st	Principle and procedures of estimation		
9th		Principle and procedures of estimation	9th	ACP estimation
901	2nd	Principle and procedures of estimation	901	ACP estillation
	3rd	Reference values & Clinical significance		
	1st	Reference values & Clinical significance		
10th	2nd	Reference values & Clinical significance	10th	Serum calcium estimation
	3rd	Lipid Profile		
	1st	Formation of cholesterol		
11th	2nd	High density and low density cholesterol	11th	Serum potassium estimation
	3rd	Principles and procedures of estimation		
	1st	Principles and procedures of estimation		
12th	2nd	Reference values & Clinical significance	12th	Serum total cholesterol estimation
	3rd	Triglycerides, pronciple and procedure of estimation		
	1st	Inportance of various ratios of HDL., LDL and VLDL		Serum triglyceride estimation
13th	2nd	Inportance of various ratios of HDL., LDL and VLDL	13th	
	3rd	Inportance of various ratios of HDL., LDL and VLDL		
	1st	Inportance of various ratios of HDL., LDL and VLDL		
14th	2nd	Inportance of various ratios of HDL., LDL and VLDL	14th	Estimation of HDL
	3rd	Sessional,Test		
	1st	Sessional,Test		
15th	2nd	Sessional,Test	15th	Estimation of LDL and VLDL
	3rd	Sessional,Test		
	1st	Sessional,Test		
16th	2nd		16th	Estimation of LDL and VLDL
	3rd			

LESSON PLAN

Name of the Faculty : Mrs. SNEHLATA

Discipline: DMLTSemester: 3rd

Subject : INDUSTRIAL / IN-HOUSE TRAINING-I

**Lesson Plan Duration**: 16 weeks(from 20 August, 2024 to 29 November, 2024)(According to Syllabus Scheme)

Work load(Lecture/Practical)per week(in hours): Practicals-02(hr)

Week		Theory		Practical	
	Lecture Day	Topic(including assigment/test)	Practical Day	Topic	
	1st				
1st	2nd		1st	Report writing based on industrial training.	
	3rd				
	1st				
2nd	2nd		2nd	Report writing based on industrial training.	
	3rd				
	1st			Preparation of Power Point Slides based on industrial training and presentation by the	
3rd	2nd		3rd	candidate.	
	3rd				
	1st			Preparation of Power Point Slides based on industrial training and presentation by the candidate.	
4th	2nd		4th		
	3rd				
	1st			Preparation of Power Point Slides based on industrial training and presentation by the	
5th	2nd		5th	candidate.	
	3rd				
	1st			Internal Evaluation based on quality of Report, PPT preparation, PPT presentation and	
6th	2nd		6th	answer to queries	
	3rd			unswer to queries	
	1st			Internal Evaluation based on quality of Report, PPT preparation, PPT presentation and	
7th	2nd		7th	answer to queries	
	3rd			answer to queries	
	1st			Internal Evaluation based on quality of Report, PPT preparation, PPT presentation and	
8th	2nd		8th	answer to queries	
	3rd			answer to queries	

	1st		External Evaluation based on quality of Report, PPT preparation, PPT presentation and
9th	2nd	9th	answer to queries.
	3rd		unswer to queries.
	1st		External Evaluation based on quality of Report, PPT preparation, PPT presentation and
10th	2nd	10th	
	3rd		answer to queries.
	1st		Estample Control based on soulity of Donard DDT association DDT association and
11th	2nd	11th	External Evaluation based on quality of Report, PPT preparation, PPT presentation and
	3rd		answer to queries.
	1st		External Evaluation based on quality of Report, PPT preparation, PPT presentation and answer to queries.
12th	2nd	12th	
	3rd	1	
	1st		
13th	2nd	13th	
	3rd	1	
	1st		
14th	2nd	14th	
	3rd		
	1st		
15th	2nd	15th	
	3rd	1	
	1st		
16th	2nd	16th	
	3rd	1	